



Screening **purple bacteria** for their growth kinetics on **volatile fatty acids**: paving the way for efficient production of **edible biomass** on fermented waste

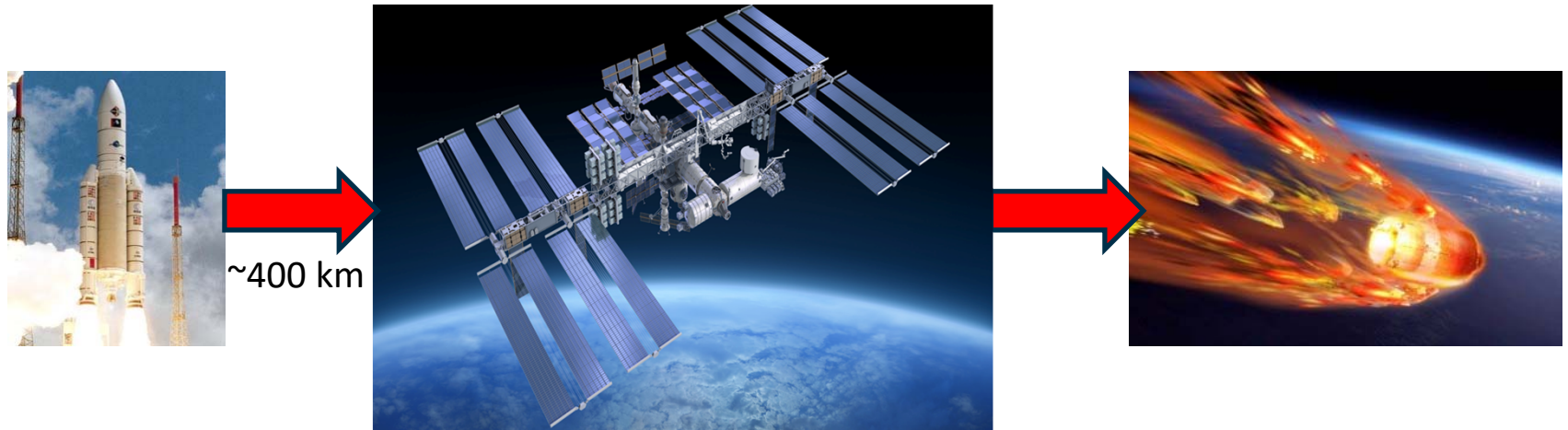
**Siegfried E. Vlaeminck & Abbas Alloul**



Sustainable Energy,  
Air & Water Technology  
University of Antwerp

# Life support in Space: from linear to circular

TODAY



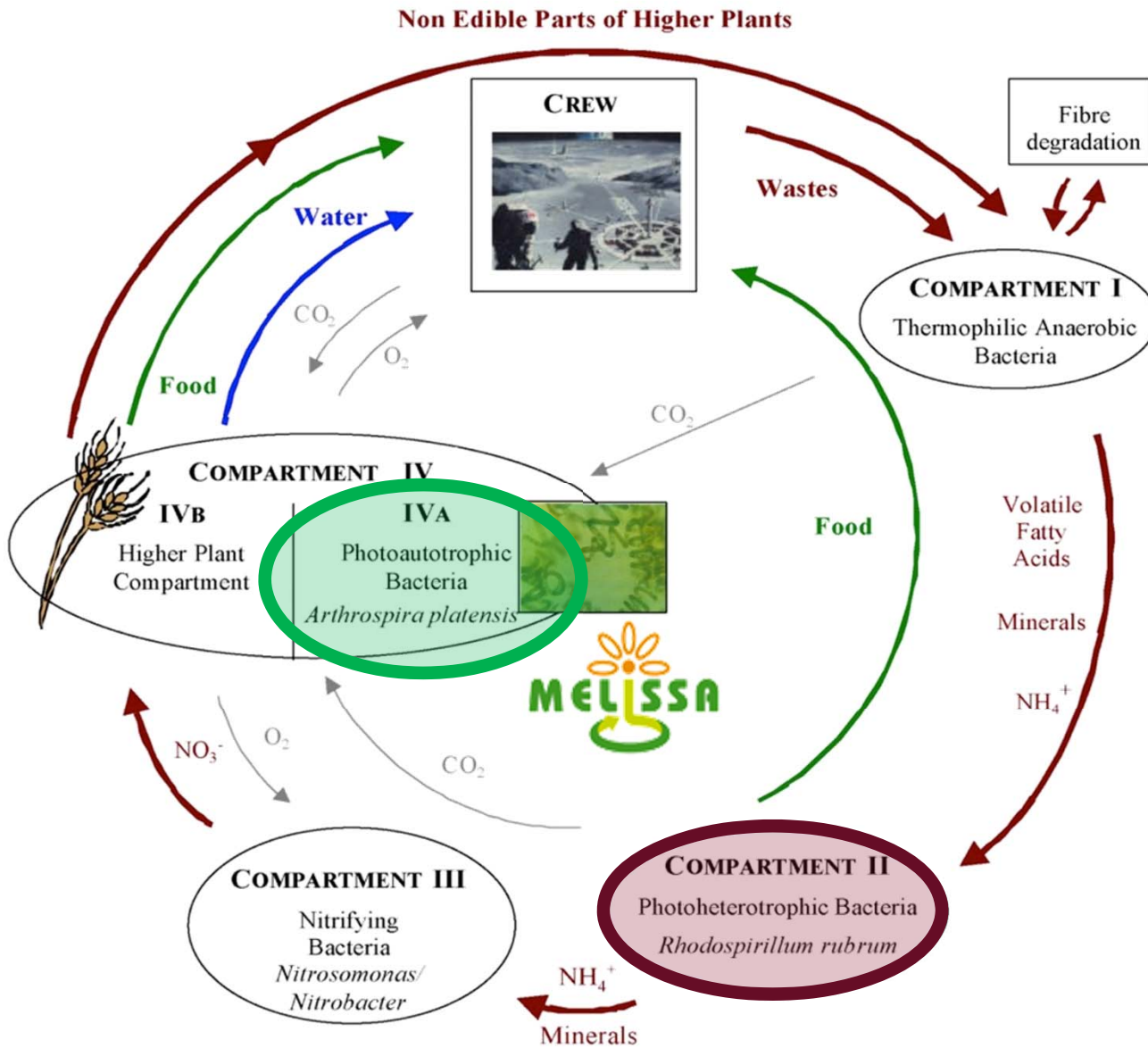
TOMORROW



(modified after Christophe Lasseur)



# MELiSSA Cycle



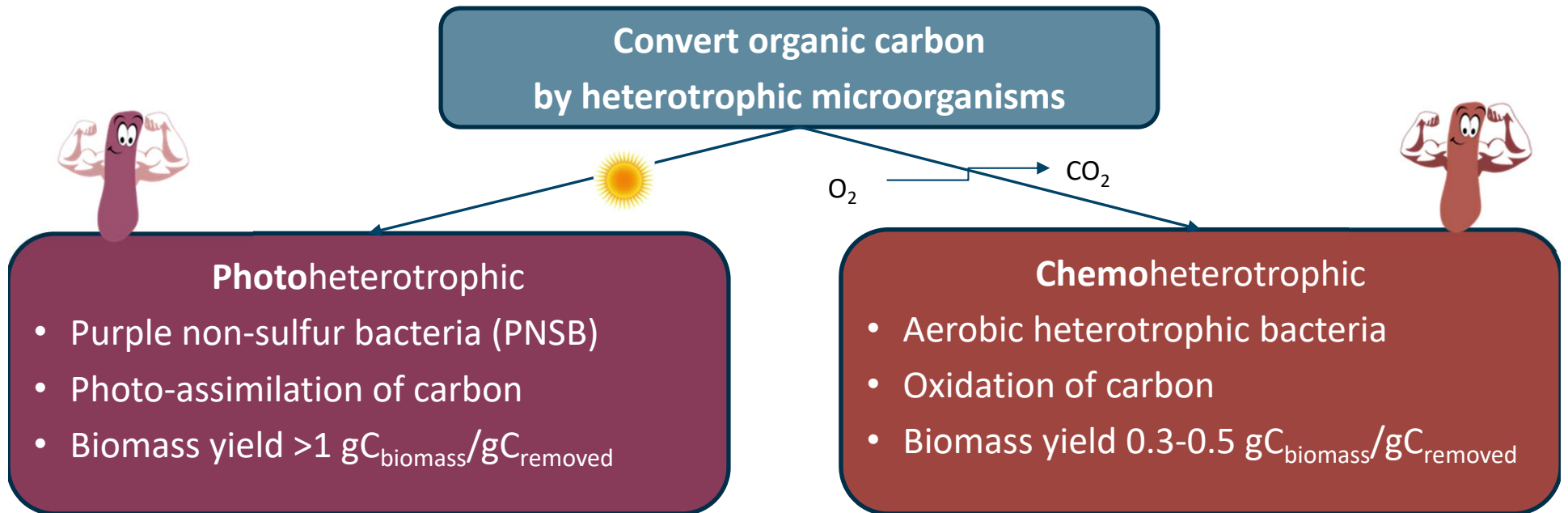
## Food production:

- Plants
- Microbes: single-cell protein
  - **CII: heterotrophic**
  - **CIVa: autotrophic**

([http://www.esa.int/Our\\_Activities/Space\\_Engineering\\_Technology/Melissa](http://www.esa.int/Our_Activities/Space_Engineering_Technology/Melissa))

## CII: Conversion of volatile fatty acids to ...

- **Microbial biomass**
  - Single cell protein => Food for crew
- Inorganic carbon  $\text{CO}_2$



## Rhodospirillum rubrum: historic choice for CII

- *Rsp. rubrum* consumes **broad spectrum of organics**
- What about the **ALiSSE** criteria the multi-parametric approach to evaluate and compare RLSS:
  - **High efficiency**
  - **Low mass**
  - **Low energy**
  - **High safety**
  - **Few crew time**

Carbon sources:	Alphaproteobacteria														Betaproteobacteria				
	Rhodospirillales					Rhizobiales					Rhodobacterales				Rhodocyclales and Burkholderiales				
	<i>Rhodospirillum rubrum</i>	<i>Rhodospirillum photometricum</i>	<i>Phaeospirillum fulvum</i>	<i>Rhodospira globiformis</i>	<i>Rosaspina medusaeina</i>	<i>Rhodomicrobium vannielii</i>	<i>Rhodobium orientis</i>	<i>Rhodospirillum roseus</i>	<i>Rhodospseudomonas palustris</i>	<i>Rhodospirillum rubrum</i>	<i>Rhodospirillum rubrum</i>	<i>Rhodospirillum rubrum</i>	<i>Rhodospirillum rubrum</i>	<i>Rhodospirillum rubrum</i>	<i>Rhodospirillum rubrum</i>	<i>Rhodospirillum rubrum</i>	<i>Rhodospirillum rubrum</i>	<i>Rhodospirillum rubrum</i>	<i>Rhodospirillum rubrum</i>
Acetate	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Aspartate	±	±	-	+	-	nd	-	±	+	-	nd	±	nd	nd	nd	+	-	+	+
Benzoate	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	+	-	-	-
Butyrate	+	+	-	+	+	+	+	+	±	-	+	+	+	+	+	+	±	+	+
Caproate	+	+	-	nd	+	+	+	+	-	±	-	+	nd	nd	+	+	+	nd	-
Caprylate	nd	-	-	-	+	-	+	+	-	-	nd	+	-	nd	+	+	-	±	nd
Citrate	-	-	-	-	-	-	±	-	±	-	-	±	nd	+	+	-	-	+	-
Ethanol	+	+	+	-	+	-	-	±	-	+	+	-	nd	-	+	-	-	±	±
Formate	-	-	nd	-	±	-	-	+	±	-	-	+	+	-	-	-	-	±	-
Fructose	±	+	-	+	-	+	-	±	+	-	-	+	+	+	+	-	-	+	+
Fumarate	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Glucose	-	+	±	+	-	+	-	±	+	±	±	+	+	+	+	+	+	+	+
Glutamate	+	-	nd	-	+	-	±	-	+	-	+	+	+	+	+	-	-	+	-
Glycerol	-	-	-	-	±	-	-	+	±	-	-	-	+	+	+	-	-	-	-
Lactate	+	+	-	+	+	+	+	+	nd	±	±	+	+	+	+	-	+	+	±
Malate	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Mannitol	+	-	+	-	-	±	-	±	+	-	±	±	+	+	+	-	-	-	+
Methanol	±	-	±	-	±	-	-	±	-	±	-	-	-	-	±	-	-	±	-
Propionate	+	±	+	-	+	+	-	+	+	+	+	+	+	+	+	-	±	±	-
Pyruvate	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Succinate	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Tartrate	-	-	nd	+	-	-	-	+	-	-	±	-	-	-	+	-	-	±	nd
Valerate	+	+	-	-	+	+	+	+	+	+	-	+	nd	nd	+	+	-	+	+

(Bergey's Manual of Systematic Bacteriology Volume Two)

# Study objectives

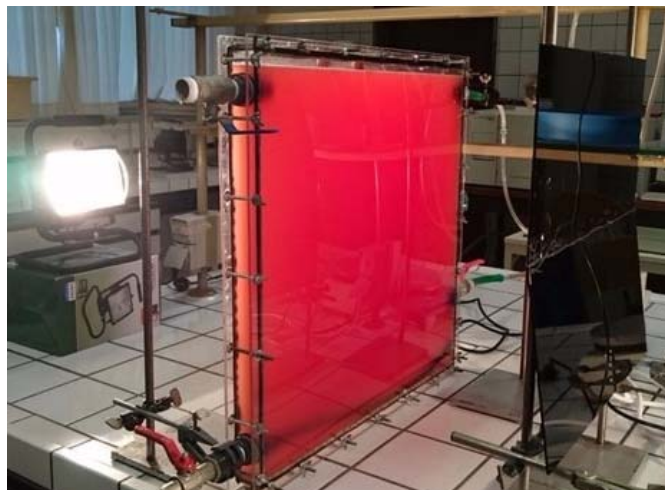
## Part 1: Batch growth

- Map the effect of VFA on growth rate and uptake profile for purple bacteria
- Select strain or community for microbial protein production

## Part 2: Continuous growth

- Optimize operational conditions of a photobioreactor to maximize protein production

## Part 3: Terrestrial valorization of PNSB



# Batch growth: Experimental variations

Carbon source: Volatile fatty acids (VFA)

1. C2: Acetic acid
2. C3: Propionic acid
3. C4: Butyric acid
4. C2/C3/C4: VFA mix (1/1/1 C-ratio)

Purple bacteria species/communities

1. *Rhodospirillum rubrum*
2. *Rhodopseudomonas palustris*
3. *Rhodobacter sphaeroides*
4. Synthetic community, SynC (1/1/1 VSS ratio of *Rsp. rubrum*, *Rps. palustris* and *Rba. sphaeroides*)
5. Purple bacteria enrichment community:
  - 1/1/1 VSS ratio of sewage activated sludge (Aquafin), sediment of local pond and dairy activated sludge
  - Enriched under IR light (filter) with VFA mix (1/1/1 C-ratio)

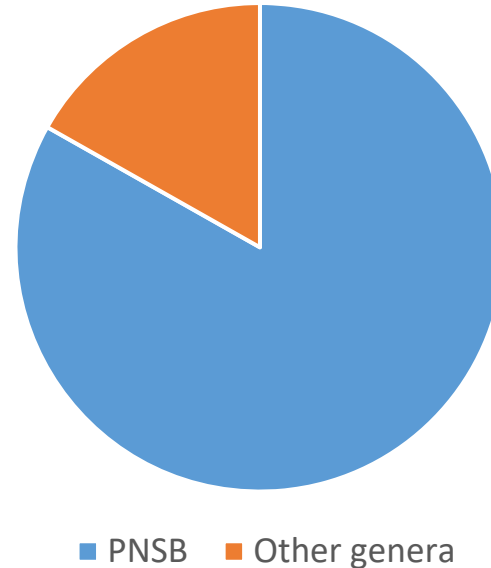


Credits to Maarten Muys UAntwerpen

## Batch growth: Microbial composition of purple bacteria enrichment Community

### MiSeq Illumina

- V4 region 16S rDNA
- Amplification 233 bp

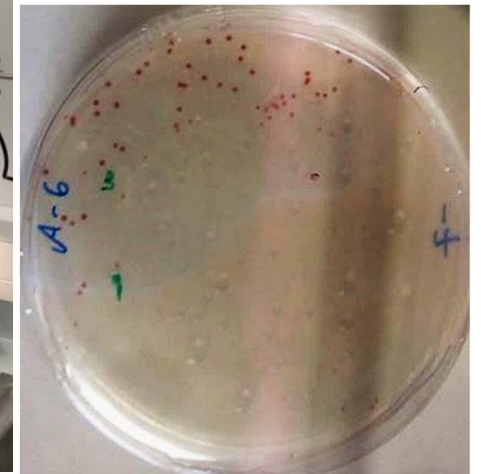
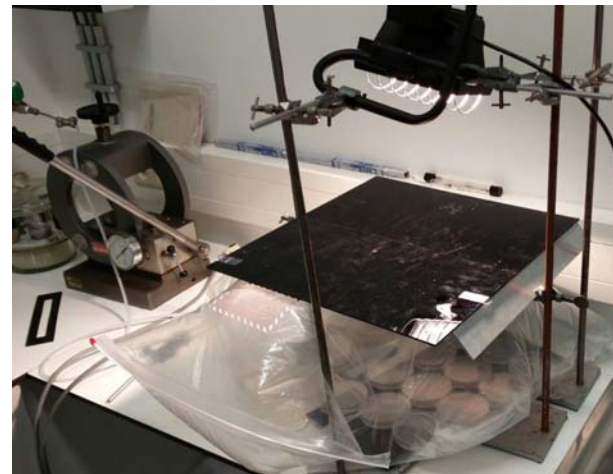


### Enrichment success!

- Enrichment conditions selective for PNSB
- OTU of PNSB dominated the microbial community

### Dilution to extinction

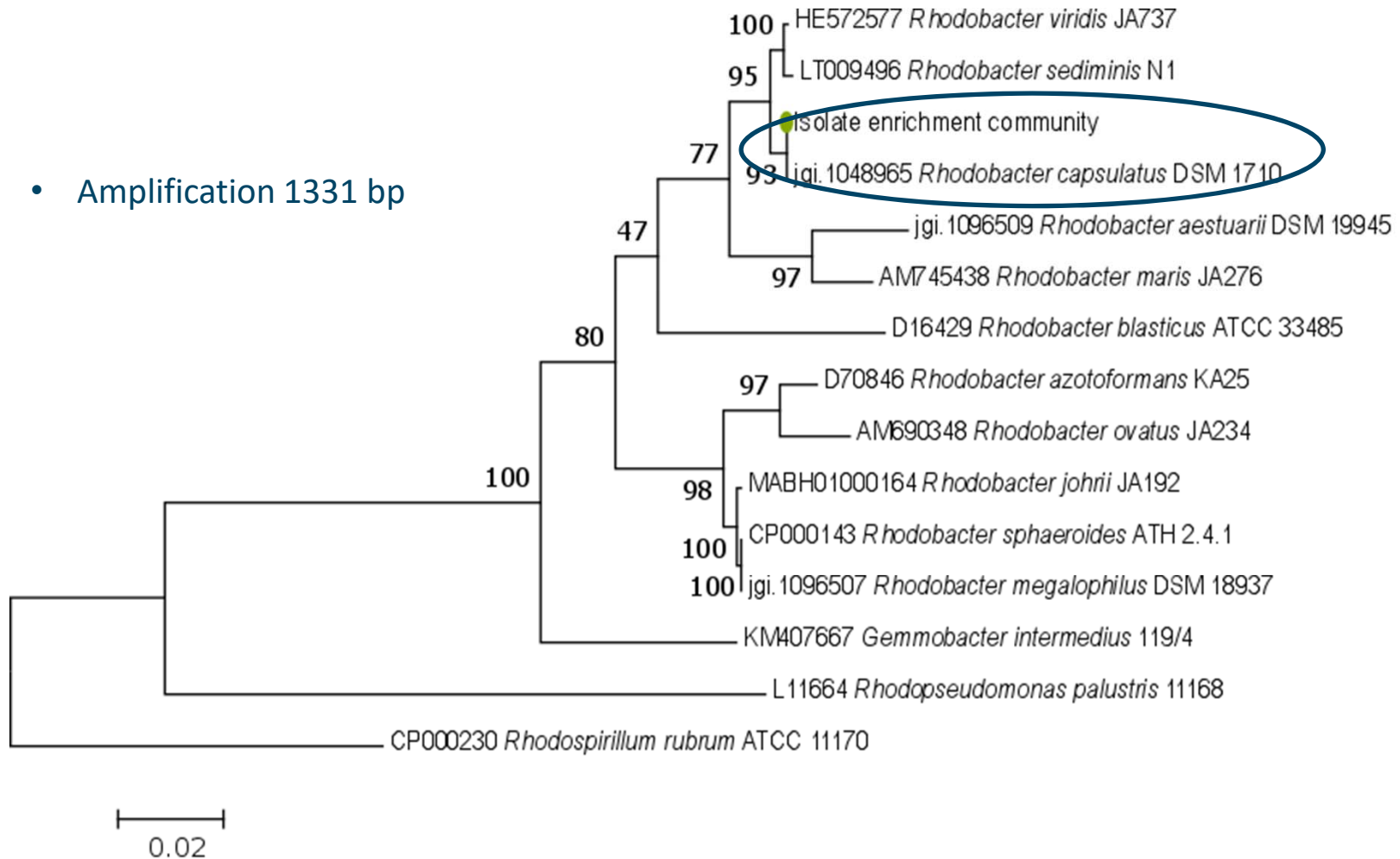
Isolation dominant species =>





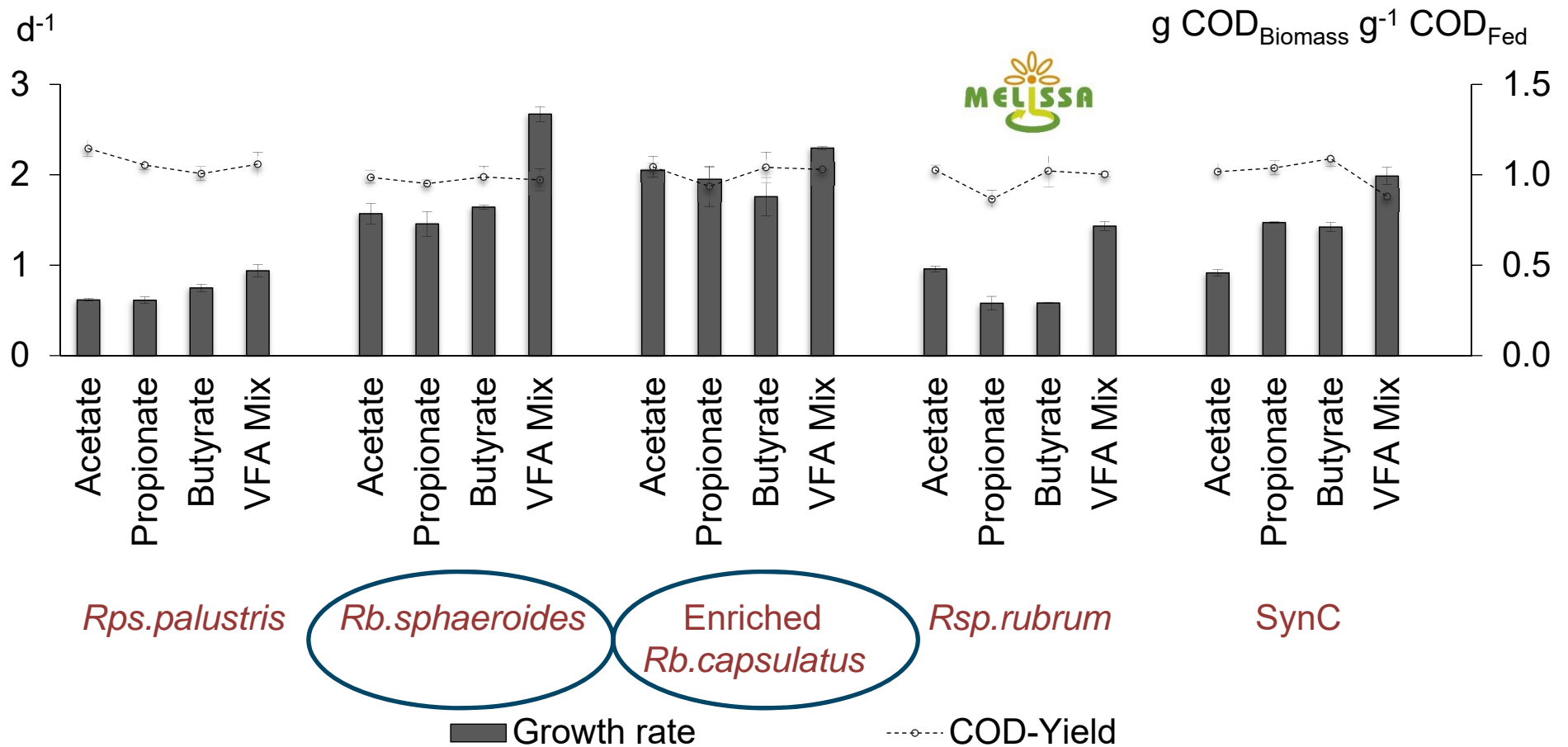
## Batch growth: Characterization of isolate - Sanger sequencing and Phylogenetic tree

- Amplification 1331 bp



***Rhodobacter capsulatus*** is most dominant species

# Batch growth: Effect of carbon on growth rate and biomass yield

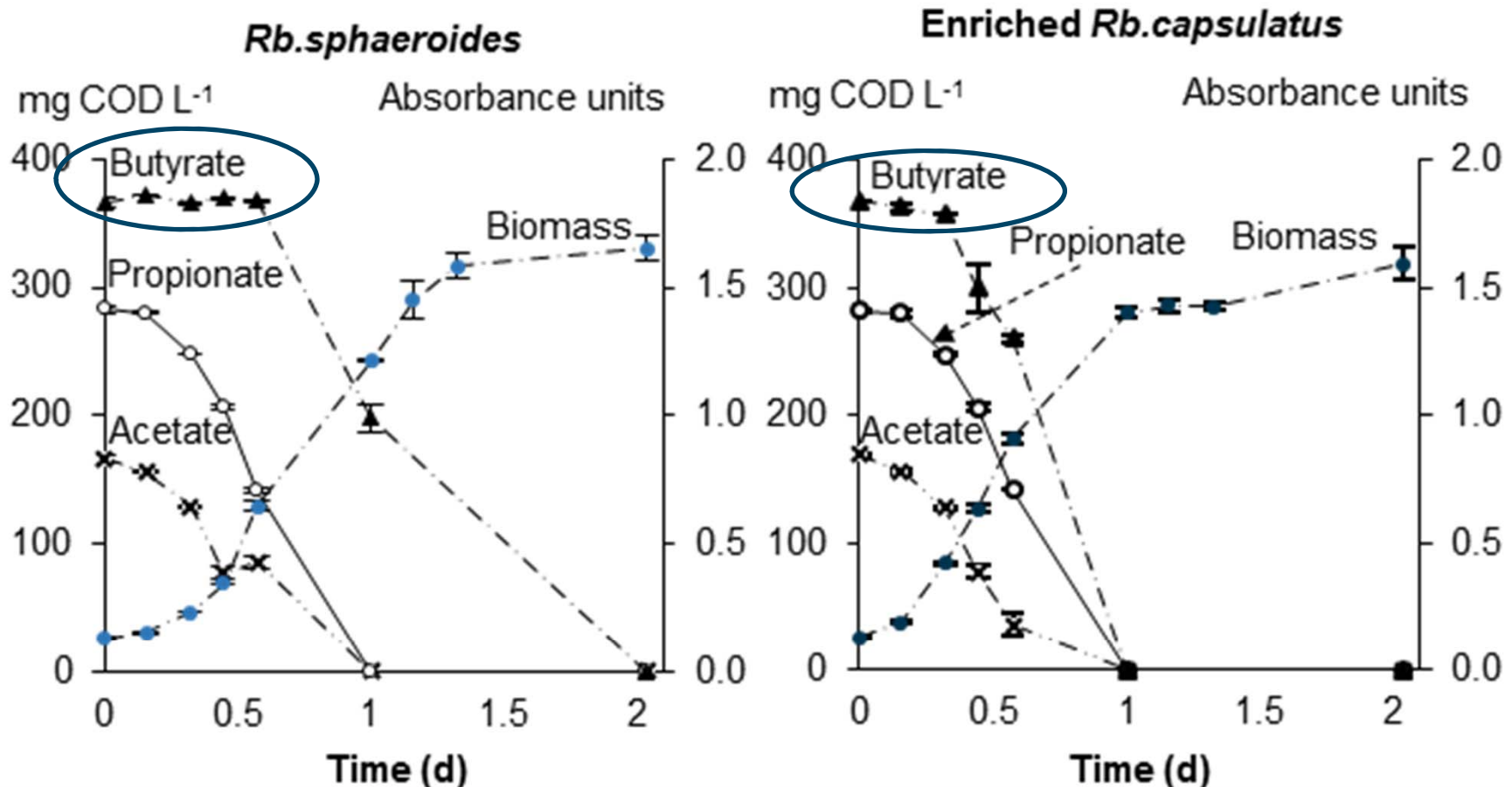


**Growth rates => Select species based growth rate vs. carbon source for MELiSSA**

- VFA mix **boosts** the growth rate for all strains/communities
- **Max  $\mu$**  for *Rba. capsulatus* enrichment community and *Rba. sphaeroides* (VFA mix)

**Full usage of COD: ~ 1 g COD to biomass/g COD removed in line with literature**

## Batch growth: Preferential carbon uptake for *Rhodobacter*



- **Lag of butyrate** for both enrichment community and *Rhodobacter sphaeroides*
- Do not overdose with synthetic medium
- Same observation for *Rsp. rubrum* experiments of De Meur (UMons, 2017)

# Study objectives

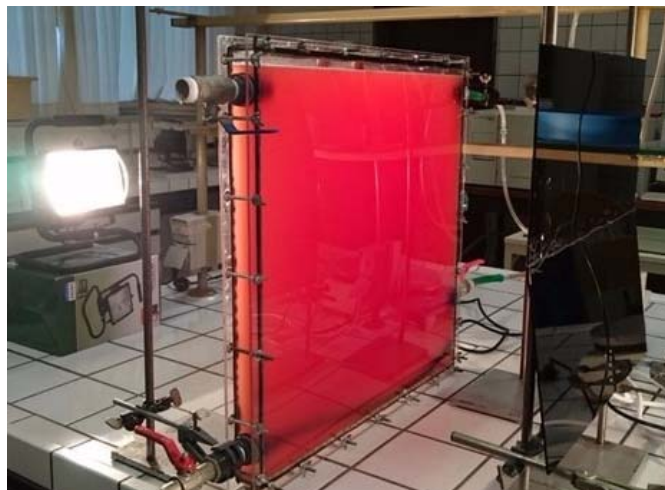
## Part 1: Batch growth

- Map the effect of VFA on growth rate and uptake profile for purple bacteria
- Select strain or community for microbial protein production

## Part 2: Continuous growth

- **Optimize operational conditions of a photobioreactor to maximize protein production**

## Part 3: Terrestrial valorization of PNSB



# Continuous growth: Operation of a photobioreactor

## Goal

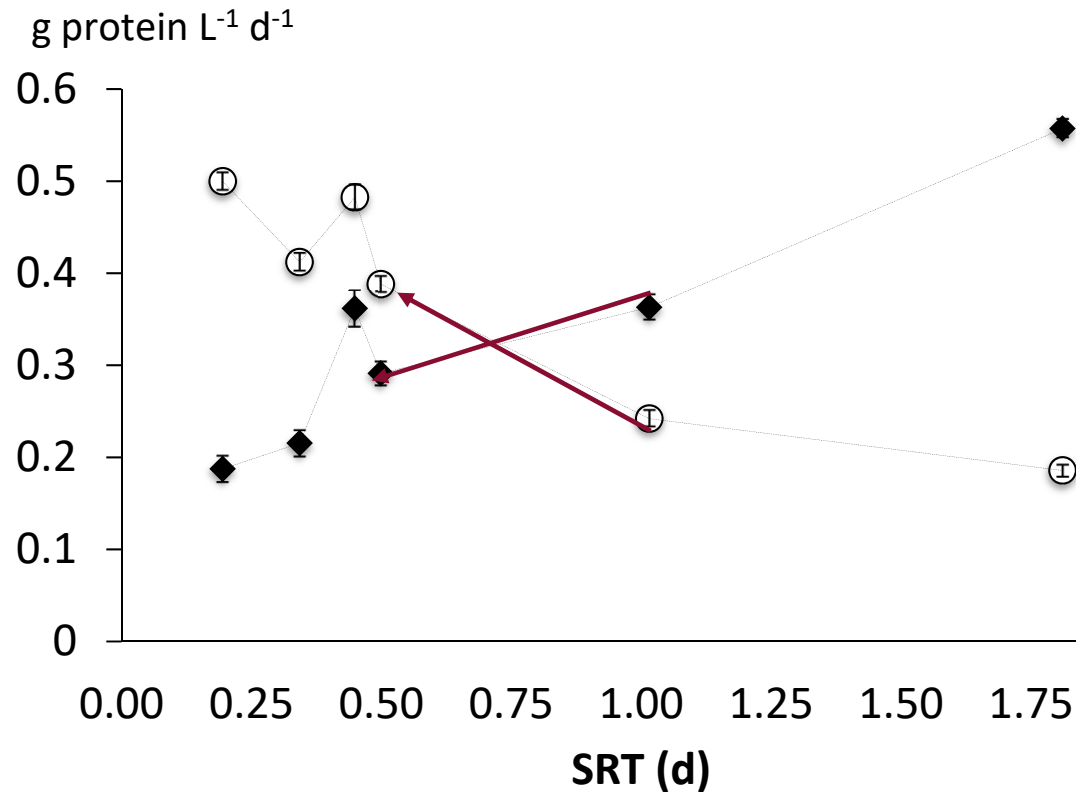
- Optimize operational conditions of a photobioreactor to maximize protein production
- Operate reactor at 6 different SRT 2 d, 1 d, 0.5 d (2 times) , 0.38 d and 0.25 d

$$SRT(d) = \frac{\text{Volume reactor (m}^3\text{)}}{\text{Flow rate (}\frac{\text{m}^3}{\text{d}}\text{)}}$$

## Hypotheses

- Shorter SRT
  - Will increase the protein productivity
  - Result in a more dominant PNSB community

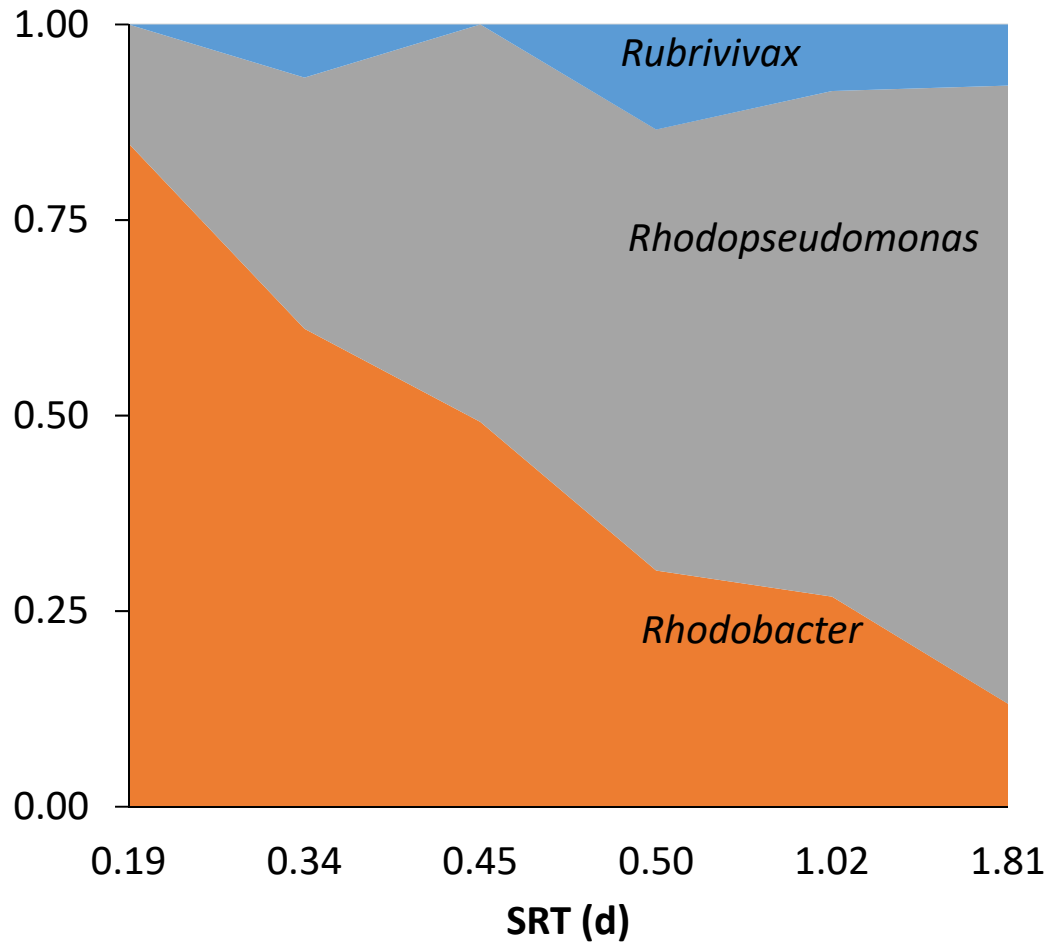
# Continuous growth: Effect of sludge retention time on protein productivity and protein concentration



○ Protein productivity

- Protein **productivity** increased with shorter SRT (no clock form observed), yet **trade of between** harvesting => **economic** analysis required

# Continuous growth: Change in microbial community during reactor operation



- *Rhodospseudomonas* dominant at long SRT
- *Rhodobacter* at short SRT

# Study objectives

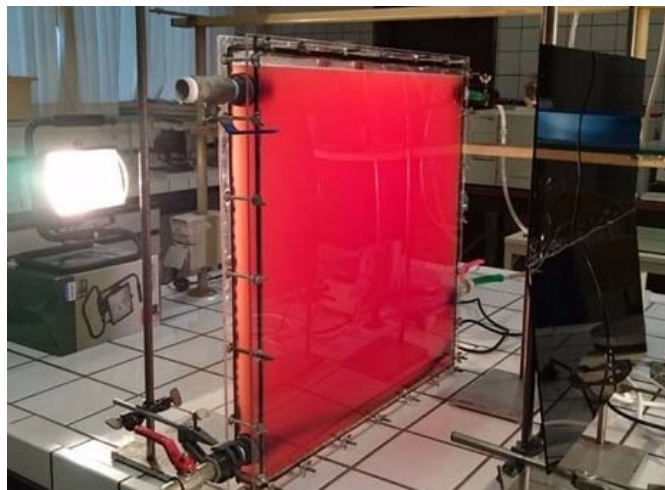
## Part 1: Batch growth

- Map the effect of VFA on growth rate and uptake profile for purple bacteria
- Select strain or community for microbial protein production

## Part 2: Continuous growth

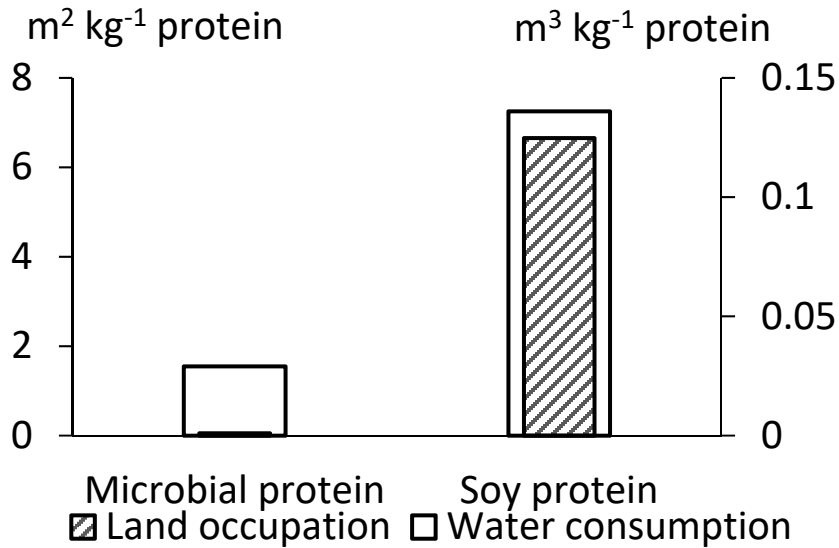
- Optimize operational conditions of a photobioreactor to maximize protein production

## Part 3: Terrestrial valorization of PNSB

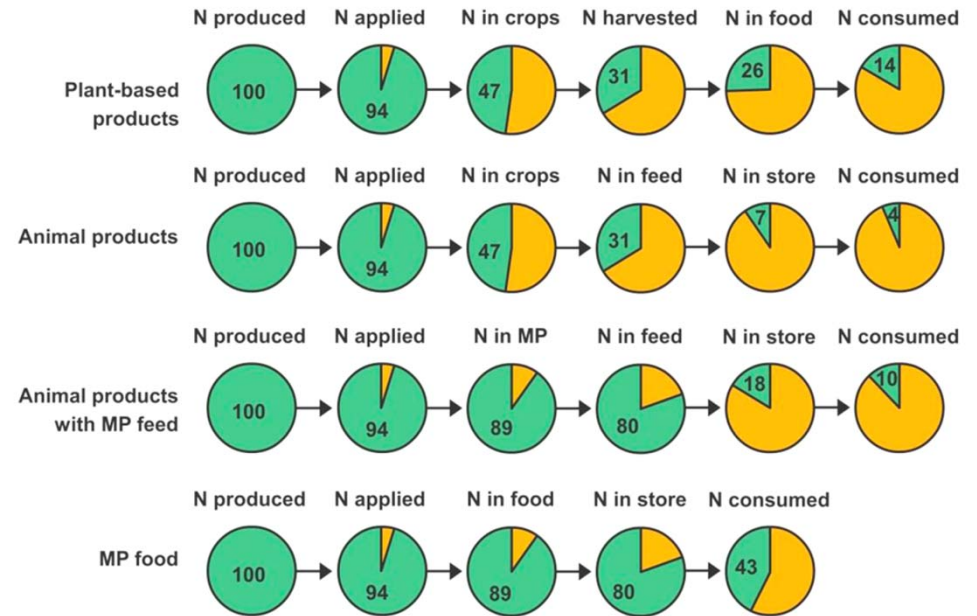




# Sustainability advantages of microbial protein (MP): Water, space and fertilizer efficiency



5X less water      128X less arable land  
(based on Matassa et al., 2016)



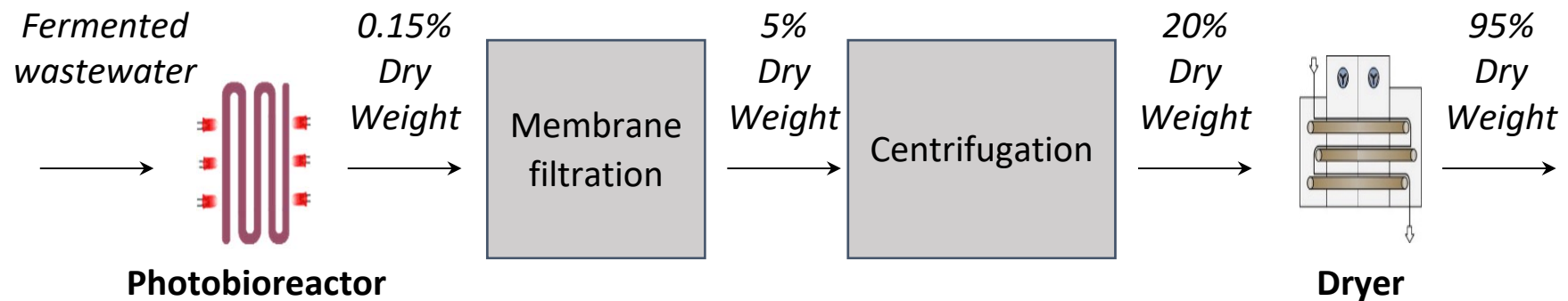
2.5-11X more nutrient-efficient than animal based products  
(Pikaar et al., 2017)



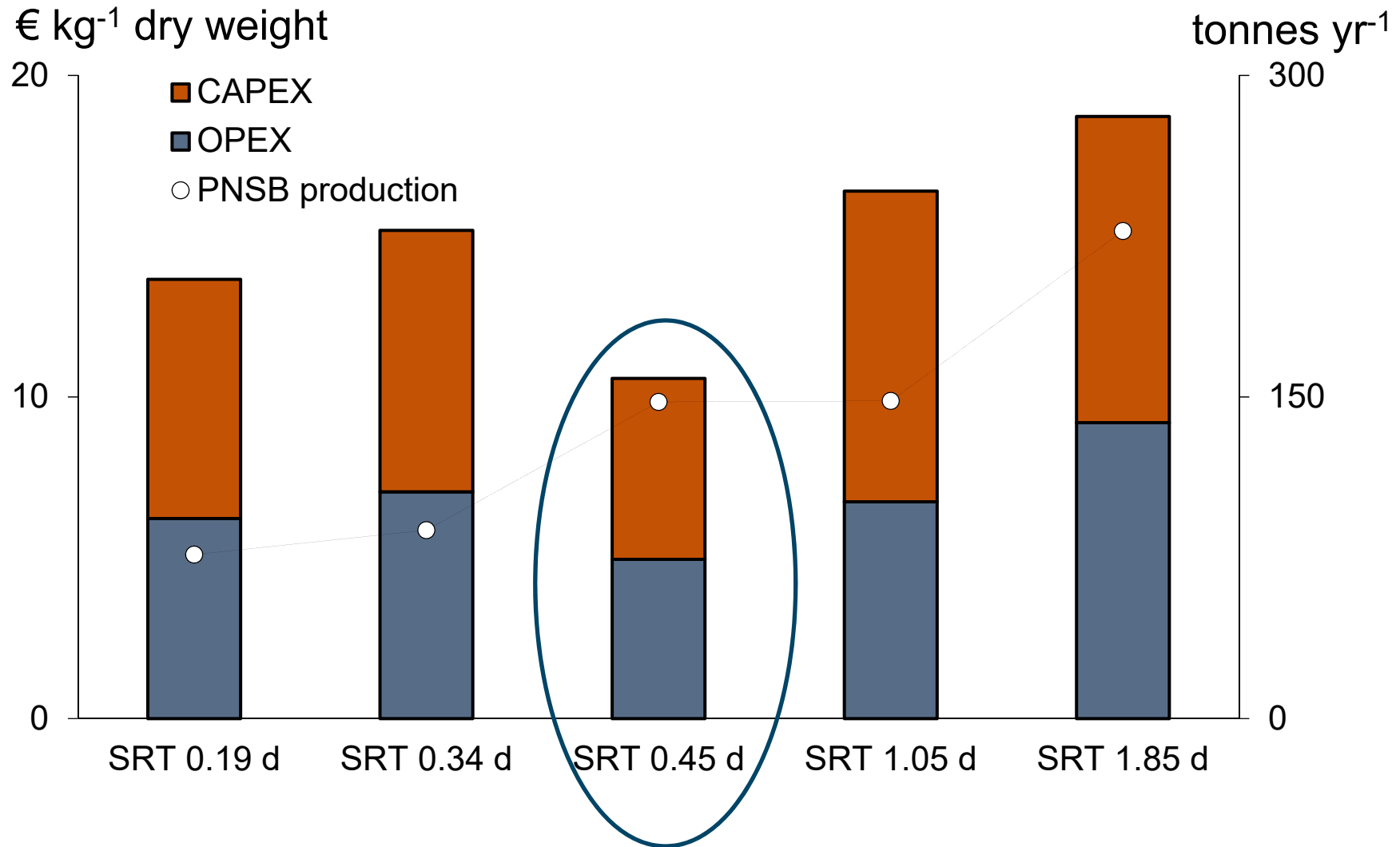
# Terrestrial valorization: Economic assessment for PNSB

## Assumptions

- Fermented brewery wastewater
- Tubular photobioreactor intermittent (1sec on/ 1sec off) with infrared LED 810 nm
- Variables: SRT and final dry weight concentration reactor (results experiments)

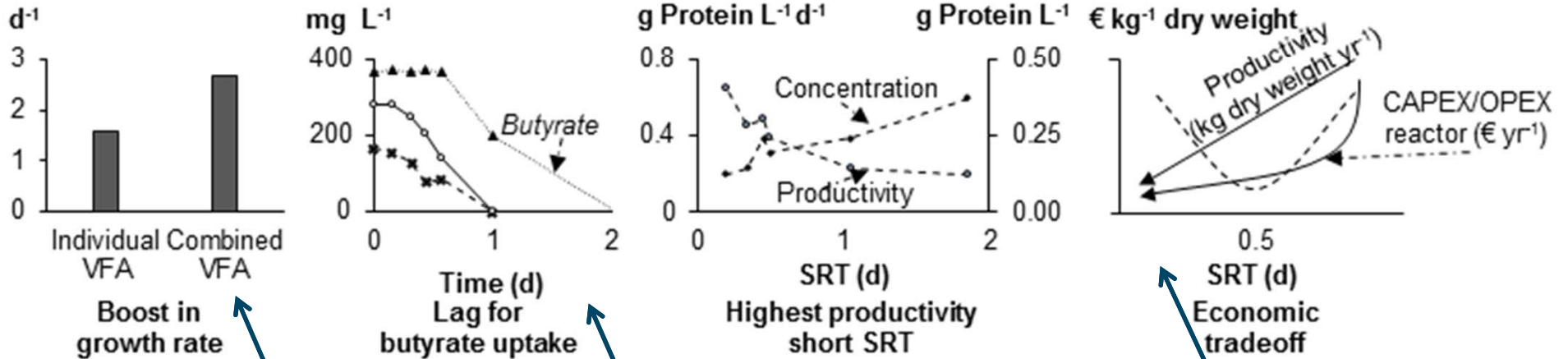
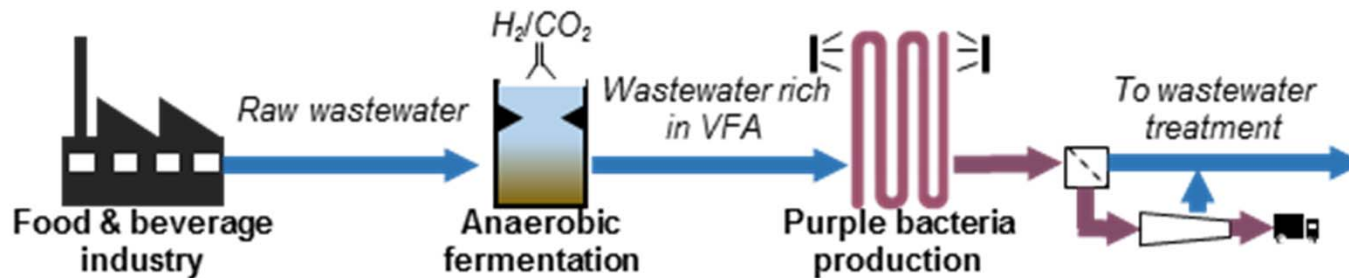


# Tradeoff between production cost and biomass production



**SRT 0.50 day best option for protein production => 10 euro/kg dry weight**

# New opportunities for purple protein production on fermented waste(water)



Propionate is the preferential C source

Economically optimal conditions: SRT of 0.50 days  
=> 10 euro kg<sup>-1</sup> dry weight

More similar to realistic fermentate

Fastest conversions obtained with *Rhodobacter* spp., not *Rhodospirillum rubrum*



Microbial nutrients on demand:  
 Waste -> 3 Types of **microbes** -> Fertilizer

- Resources: COD, N, P in **waste and side streams**
- Recovery: Grow **three types of micro-organisms**
- Reuse: Dried microbial biomass is **as effective as commercial organic fertilizer** at the level of:
  - Mineralization
  - Fertilization: parsley, ryegrass, tomato, surfinia

-> Promising as **sustainable next-generation fertilizers (NGF)**



Purple non-sulfur bacteria;  
*Rhodobacter sphaeroides*



Consortium of aerobic bacteria



Microalgae; Spirulina





Ir. Sander Wuyts  
(UAntwerpen)



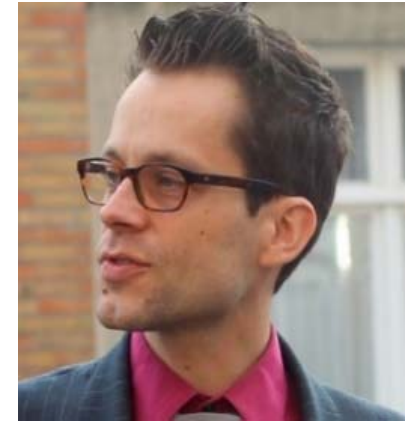
Prof. Sarah Lebeer  
(UAntwerpen)



Prof. Ilse Smets  
(KU Leuven)



Kenneth Simoens  
(KU Leuven)



[Abbas.Alloul@UAntwerpen.be](mailto:Abbas.Alloul@UAntwerpen.be)

[Siegfried.Vlaeminck@UAntwerpen.be](mailto:Siegfried.Vlaeminck@UAntwerpen.be)

## Acknowledgments

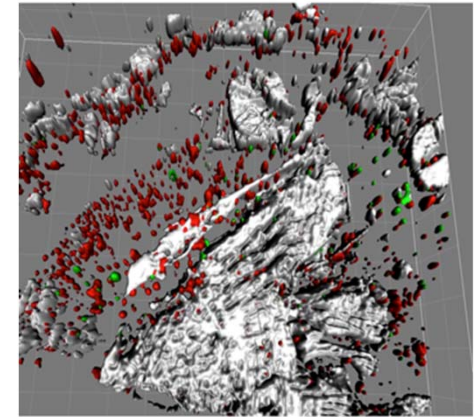


Last scientific presentation: tomorrow **14h25** in Sala **Marconi**



**Variability in nutritional value and safety of *Arthrospira* and *Chlorella* biomass necessitates smart production of microalgae for human spaceflight**

**Siegfried E. Vlaeminck, Sui Yixing, Pieter Vermeir & Maarten Muys**



- **Open access** journal (°2008)
- Editors: Kenneth N. Timmis, Juan Luis Ramos, Willem de Vos, Siegfried E. Vlaeminck, Auxiliadora Prieto
- 2016 ranking:
  - Impact factor: **3.5**
  - **Top 30%** in Microbiology (36/124) and Biotechnology & Applied Microbiology (40/158)

Environmental biotechnology - Green chemistry - Food, beverages and supplements -  
Bioenergy - Agriculture - Bioremediation - Biopolymers, biomaterials -  
Technology development - Process engineering

– AND MUCH MORE

Looking forward to receive your best work!