

CMET

Center for Microbial Ecology and Technology



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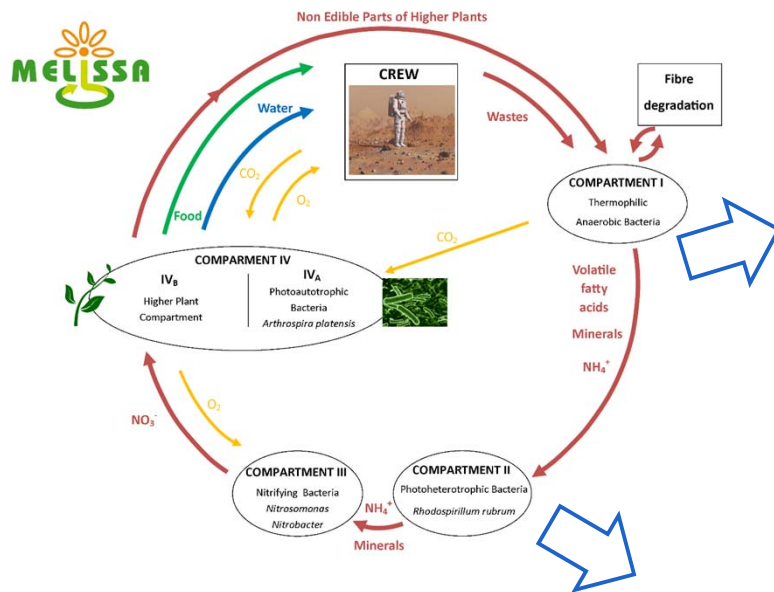
COUPLING BIOELECTROCHEMICAL OXIDATION TO C1 OF THE MELISSA LOOP

Dr. Amanda Luther 18-05-2018



MELISSA

Challenge: organics mineralization to CO₂



C1 fermentation (optimized)

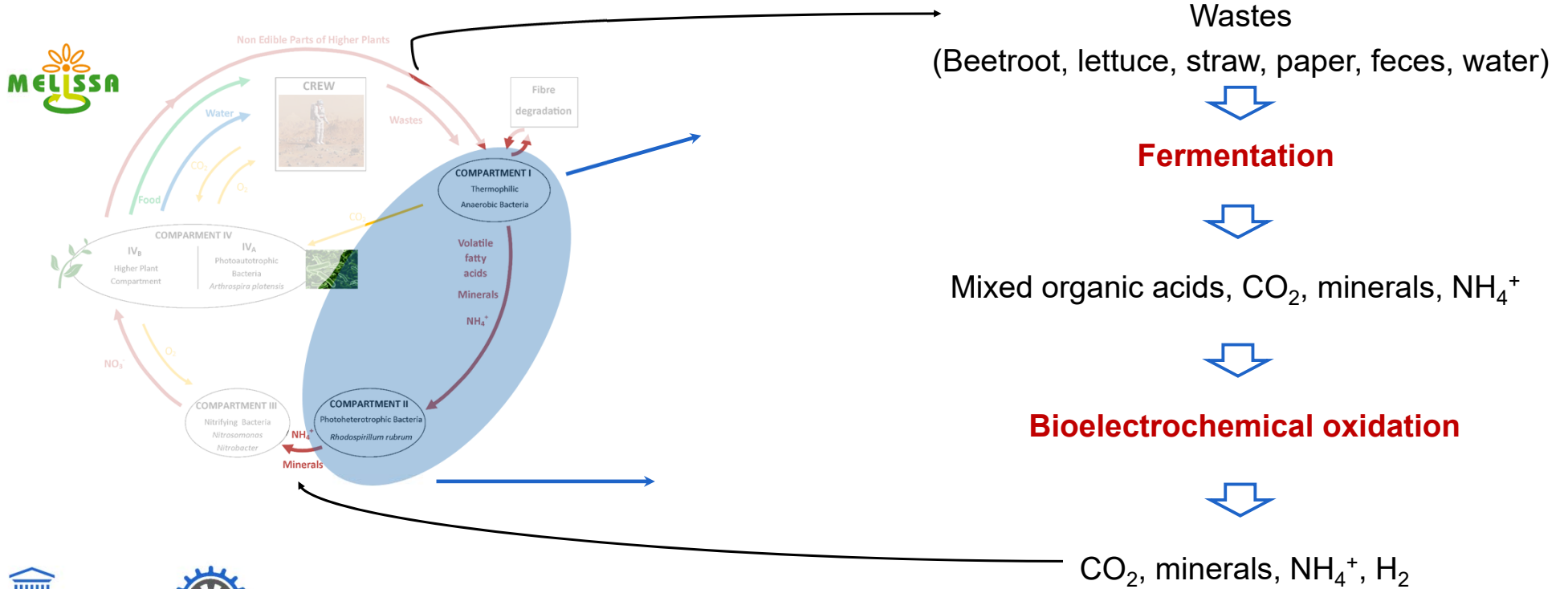
- 15% organic carbon mineralized to CO₂
- ~35% carbon (soluble) routed to C2
- ~45% of nitrogen (organics + ammonium) routed to C2

C2 compartment (aim)

- mineralize organic carbon to CO₂
- retain minerals in effluent, and nitrogen as ammonia

MELISSA

Bioanodic oxidation of fermentation products



C1 FERMENTATION

Standard MELiSSA C1

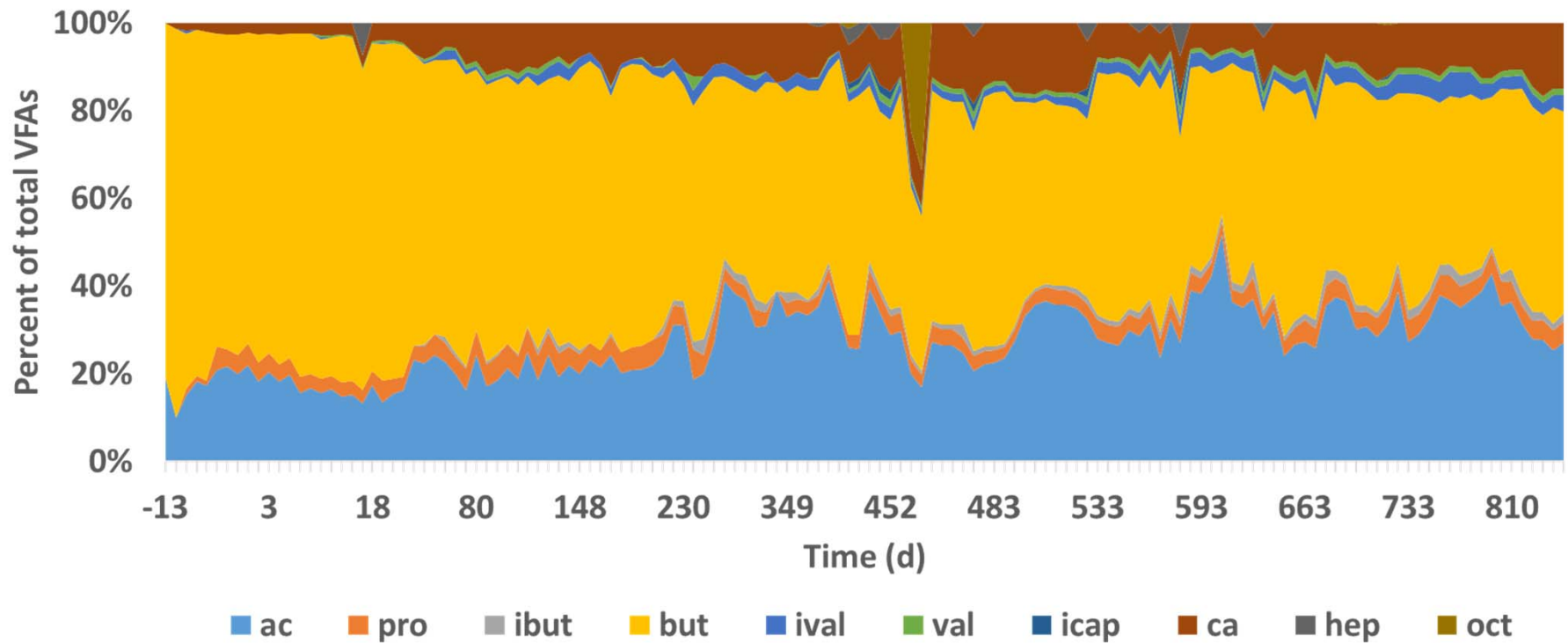
- 5 L reactor
- 55 °C
- pH 5.3
- External membrane filtration
- HRT = 10 Days
- SRT = 80 Days



INFLUENT CONTAINS REDUCED C AND N

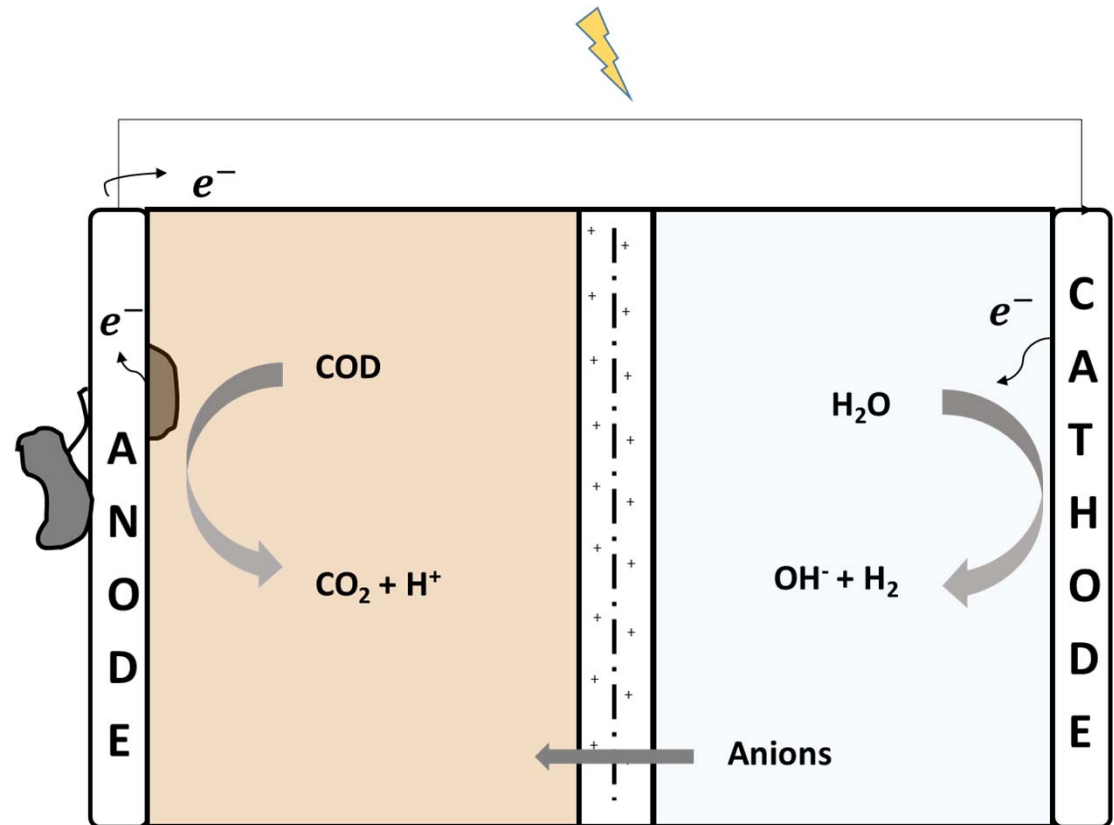
Total COD (g/L)	9.6 ± 1.6	
Total organic acids (g COD/L)	8.0 ± 0.9	(83% of total COD)
Acetic acid (g COD/L)	2.4 ± 0.1	(25% of total COD)
Butyric acid (g COD/L)	3.9 ± 0.1	(41% of total COD)
Caproic acid (g COD/L)	0.8 ± 0.0	(8% of total COD)
Ethanol (g COD/L)	1.4 ± 1.1	(15% of total COD)
Other	0.2	(2% of total COD)
Total Nitrogen (mg N/L)	270 ± 47	
Total ammonia nitrogen (mg N/L)	153 ± 23	(57% of total nitrogen)
Total organic nitrogen (mg N/L)	124 ± 57	(46% of total nitrogen)

VFA PRODUCTION IN C1



BIOANODIC OXIDATION

- Organics expressed as chemical oxygen demand (COD)
- Bio-anodic oxidation of organics produces a current
- Charge balanced by anion transfer from cathode to anode



POSSIBLE CARBON AND ELECTRON SINKS

Bio-anodic oxidation

Competing microbial
metabolism
(alternative e- acceptors)

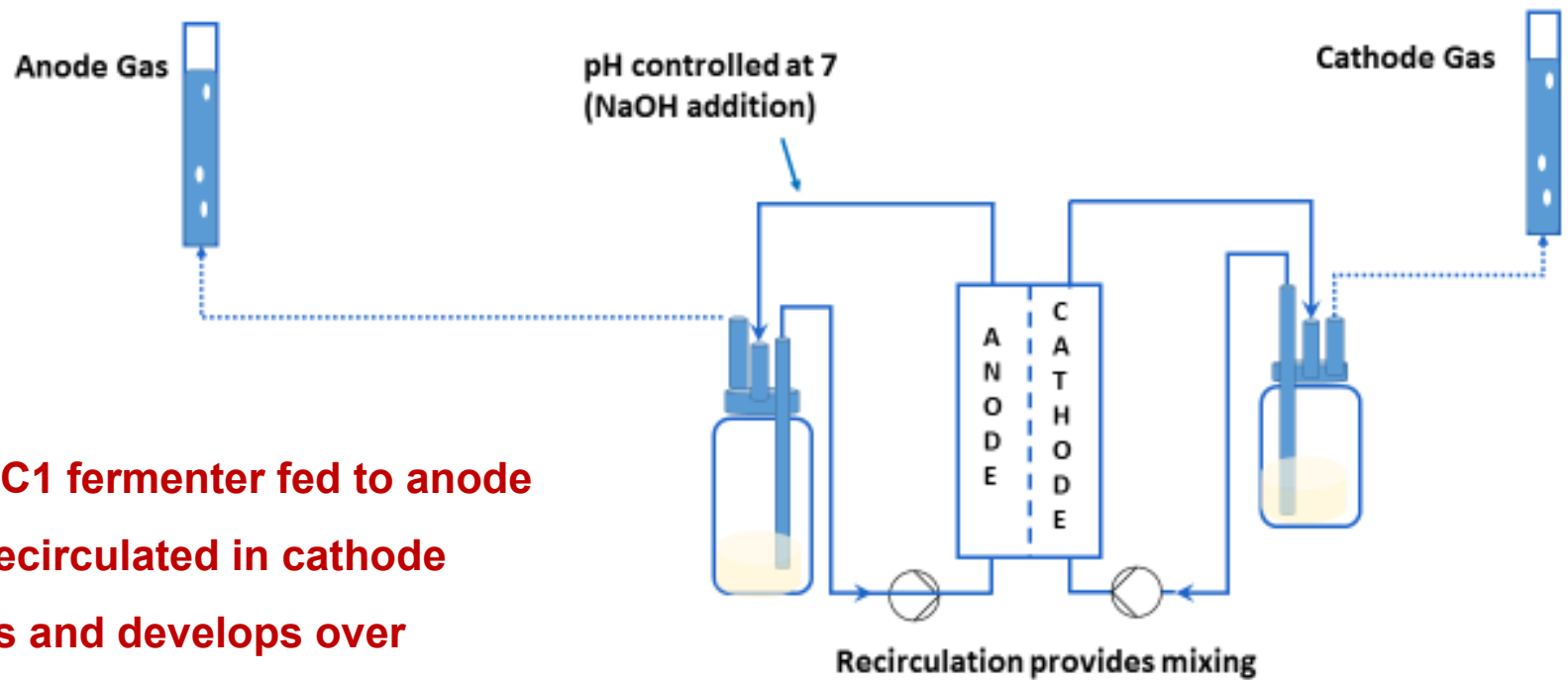
Residual substrate

Losses

Coulombic efficiency expresses the fraction of oxidized COD converted to current

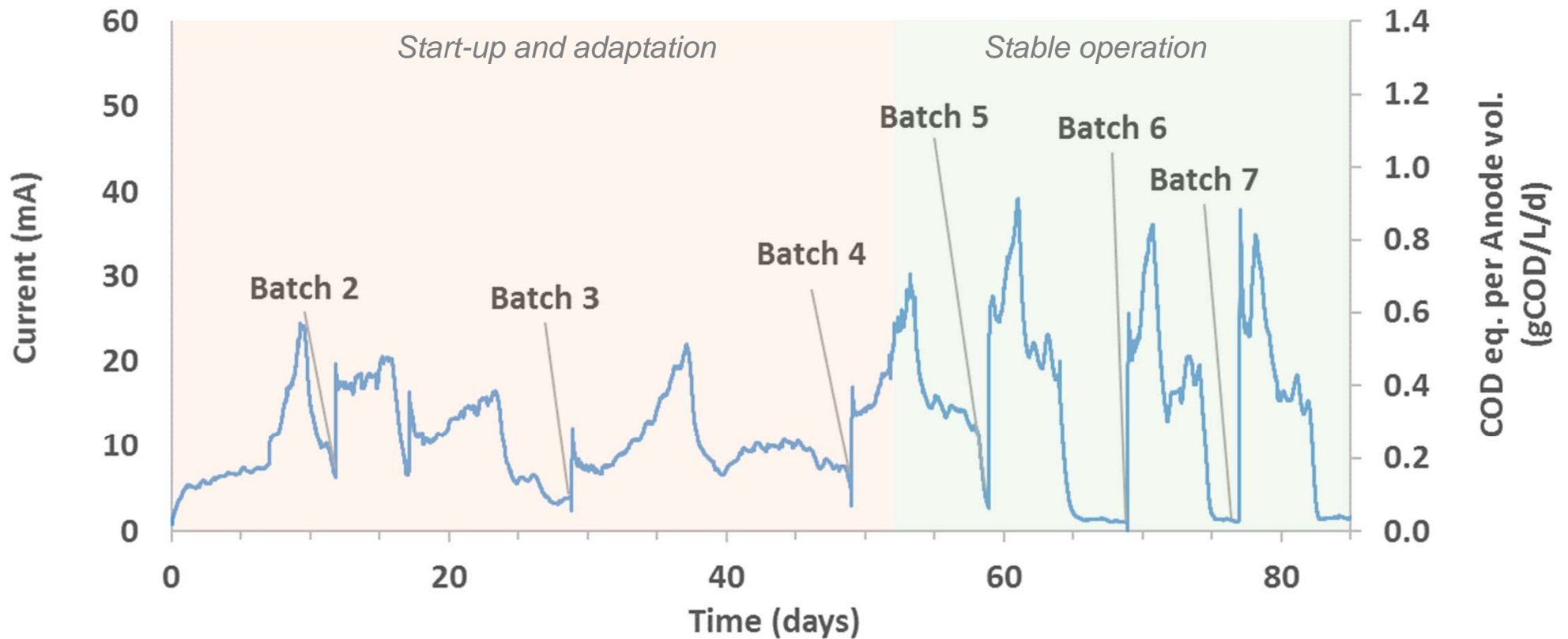
Electron balance on COD to identify sinks

BATCH TESTS ON C1 EFFLUENT



- Effluent from C1 fermenter fed to anode
- Sodium salt recirculated in cathode
- Biofilm adapts and develops over consecutive batches

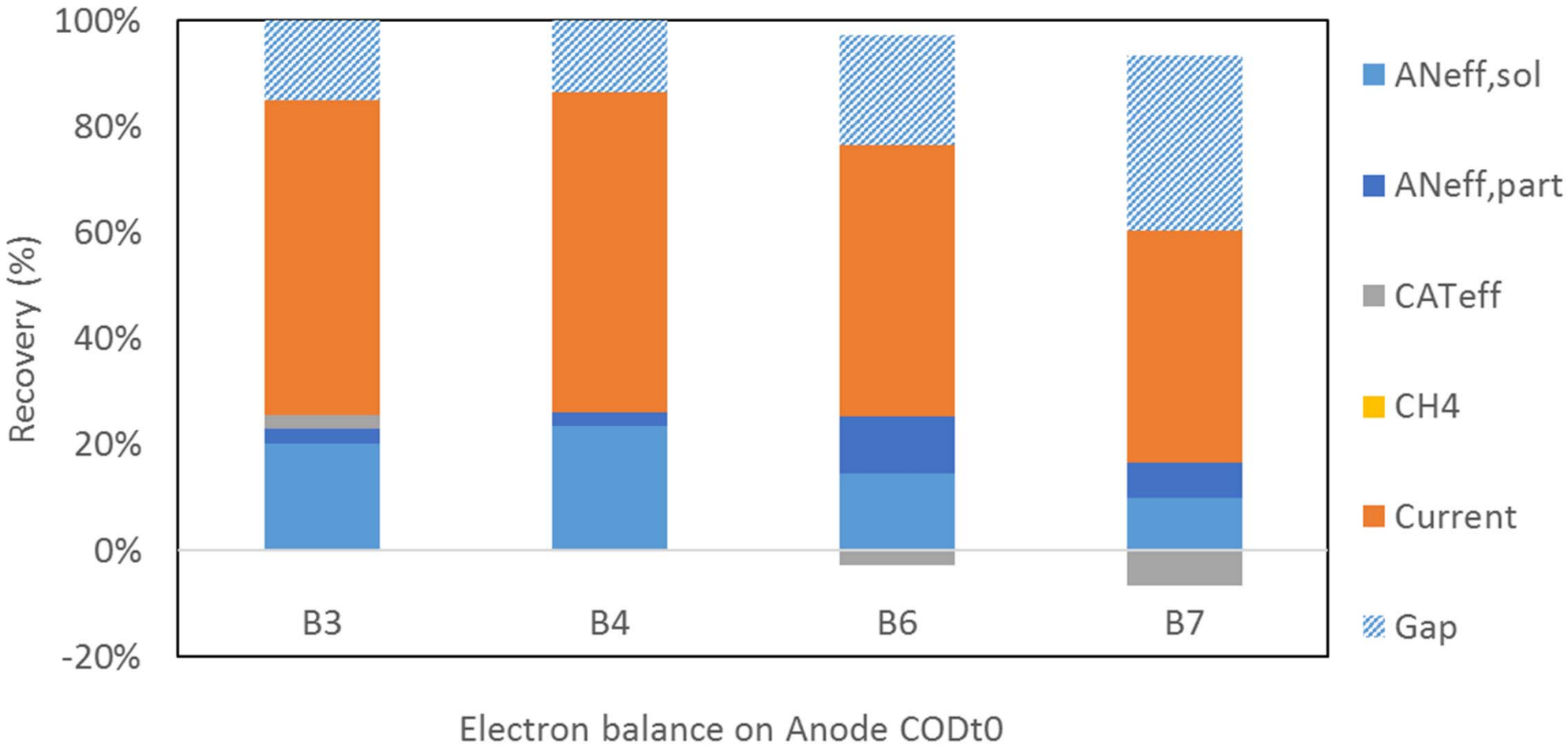
STABLE BATCH OPERATION



PERFORMANCE OVERVIEW

Batch #	RE_{COD} (%)	tVFA RE (%)	CE_{COD} (%)	pH anode (t_f)	pH cathode (t_f)
4	77	85	79	7.8	12.9
6	85	100	64	8.8	12.5
7	89	93	57	8.5	12.6

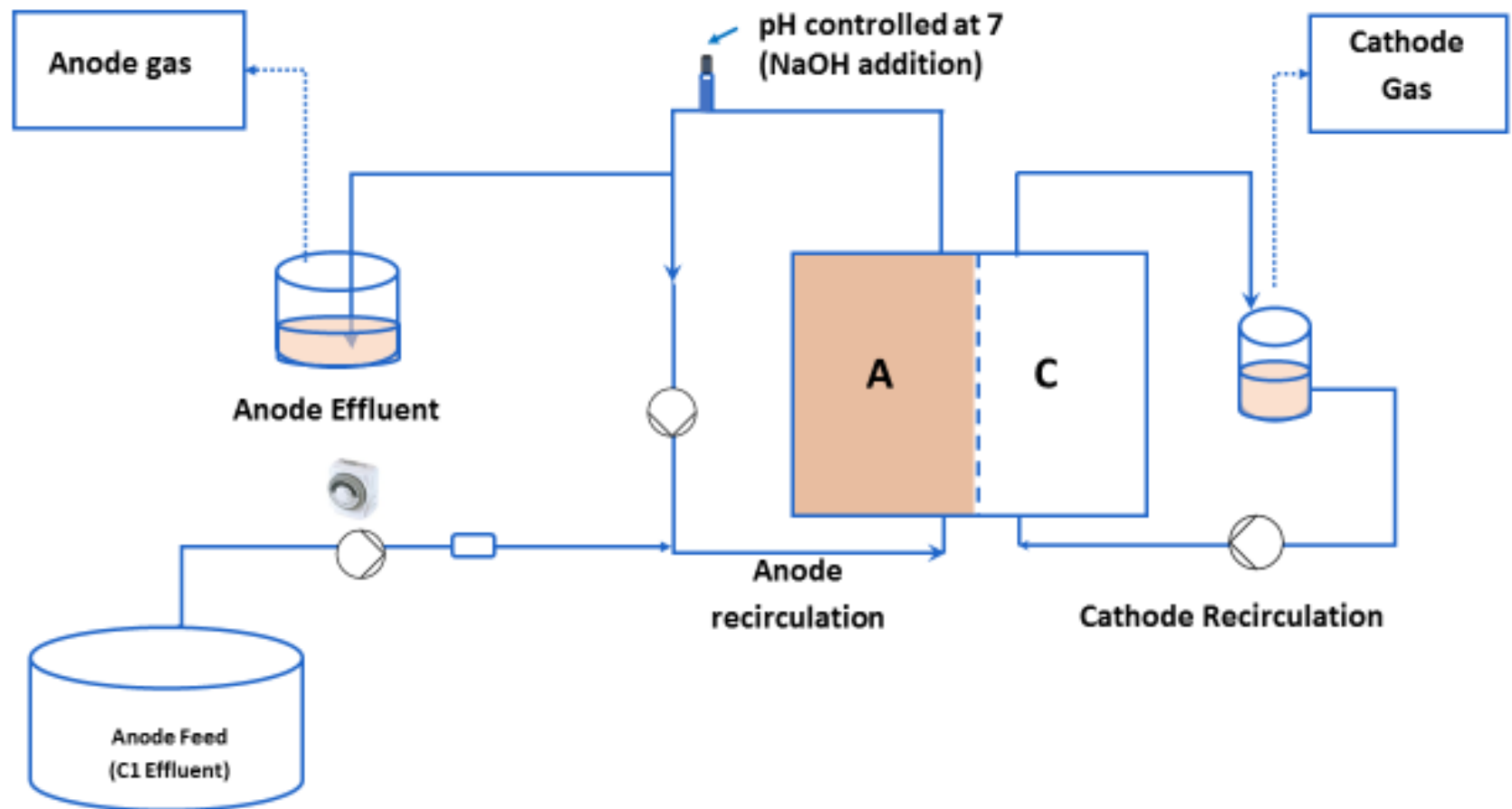
COD (ELECTRON) BALANCE



CONTINUOUS TESTS

Test 1 2 g COD L⁻¹ d⁻¹	Test 2 2 g COD L⁻¹ d⁻¹	Test 3 1 g COD L⁻¹ d⁻¹
<ul style="list-style-type: none">• Continuous feed following batch test• Same bioanode	<ul style="list-style-type: none">• modifications to set-up• Same bioanode	<ul style="list-style-type: none">• Reduced organic loading• Refresh anode

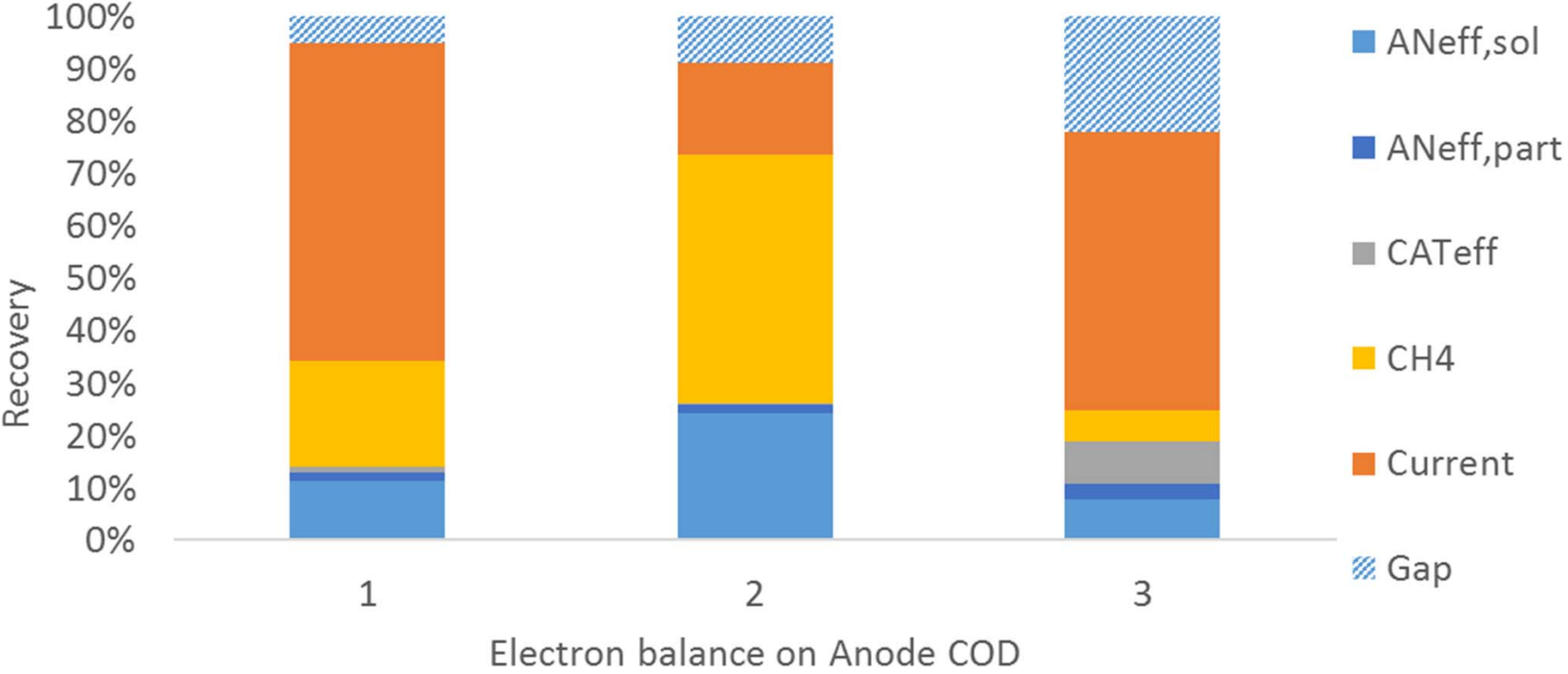
CONTINUOUS TESTS



PERFORMANCE OVERVIEW

Test #	Stable operation period (d)	COD loading ($\text{g L}_{\text{AN}}^{-1} \text{d}^{-1}$)	COD RE (%)	VFA RE (%)	CE_{COD} (%)	Anode pH	Cathode pH
1	21	2	88	96	73	7.8 ± 0.2	13.0 ± 0.1
2	63	2.4	76	82	23	7.6 ± 0.5	10.4 ± 0.6
3	33	0.7	92	98	56	7.0 ± 0.1	12.2 ± 0.4

COD (ELECTRON) BALANCE



PROOF OF CONCEPT ESTABLISHED

- Real C1 effluent is amenable to bio-anodic oxidation
- High removal efficiencies could be achieved
- Bio-anodic activity favored under batch conditions, but not sustained under continuous operation
- Limited COD removal: ~15% COD remains in effluent

NEXT STEPS

- Optimize continuous operation
- Carbon and nitrogen balance through system
- In-depth characterization of long-term performance

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GENEROUS DONORS



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